

Discipline Specific Core (200-299)



University of Kerala

Discipline	BIOTECHNOLOGY				
Course Code	UK3DSCBIT200				
Course Title	BIOMOLECULES & METABOLISM				
Type of Course	DSC				
Semester	III				
Academic Level	200 - 299.				
Course Details	Credit	Lecture per week	Tutorial per week	Practical per week	Total Hours/Week
	4	3 hours	0	2 hours	5
Pre-requisites	Fundamentals of Biochemistry, cell biology				
Course Summary	This course delves into the intricate world of biomolecules and the fundamental processes governing metabolism in living organisms. It provides an advanced understanding of the structure, function, and interplay of various biomolecules such as proteins, carbohydrates, lipids, and nucleic acids, as well as the metabolic pathways that govern energy production, biosynthesis, and cellular regulation.				

Detailed Syllabus:

Module	Unit	Content	Hrs
I	Basic constituents of Life -Overview		6
	1	Chemical Constituents of Life, Importance of water and physiological buffer system	
	2	Carbohydrates : Classification, mono, di, and polysaccharides with special	

		emphasis on starch, glycogen, cellulose, and chitin	
	3	Glycoconjugates and its biological significance. glycosides, deoxy sugars, amino sugars, sugar alcohols and sugar acids (Lectins-CS)	
II		Lipids	8
	4	Classification of lipids	
	5	Fatty Acids, Triglycerides, Phospholipids, Sphingolipids - structure, properties and reactions	
	6	Function of Steroids—cholesterol (CS) and ergosterol, phosphatidyl choline and phosphatidyl ethanolamine, cerebrosides and gangliosides	
III		Amino Acids and Proteins	10
	7	Classification of amino acids, Physical properties, Chemical reactions of amino acids	
	8	Biological significance and classification- fibrous proteins, globular proteins, conjugated proteins (CS)	
	9	Elementary study of primary, secondary, tertiary and quaternary structure of proteins; (CS-motifs) oligopeptides- glutathione; Hemoglobin- structure and functions	
	10	Nucleic acids: Base compositions, structure of purines and pyrimidines, ribose and deoxyribose, nucleoside structure, nucleotides- nomenclature, structure of polynucleotide – DNA, RNA primary structure and inter nucleotide linkage Watson and Crick double helix model of DNA, different types of RNA.	
IV		Metabolism of Biomolecules	12
	11	Metabolism basic concepts- Energy rich compounds-ATP, Common types of reaction in metabolism-Oxidation, reduction, phosphorylation, hydrolysis, hydroxylation, carboxylation. High energy compounds with structures (ATP, ADP, Creatine phosphate, 1, 3 biphosphoglycerate, PEP), role of high energy phosphate groups.	
	12	Metabolism of carbohydrates Glycolysis. Gluconeogenesis, Glycogen metabolism- glycogenesis, glycogenolysis. Regulation (Only pathway outlines, structures not required).	
	13	Metabolism of Lipids. Scheme of β - oxidation, ATP yield in β oxidation (stearate, palmitate as examples) and regulation. Basics of α - and ω -oxidation, ketone body formation, cytoplasmic system of fatty acid biosynthesis and regulation of the pathway, outline study of biosynthesis of cholesterol and bile acids (Only pathway outlines, structures not required).	
	14	Metabolism of amino acids. Reactions involved in the metabolism of amino acids- deamination, transamination and decarboxylation; coenzymes involved in these reactions. Urea cycle (structure not required). Metabolism of Nucleic Acids- De Novo & Salvage Pathway.	
V		Enzymes	9

15	Enzymes - Classification Units of enzyme activity, progress curve, effect of enzyme concentration, substrate concentration- (Michaelis-Menten equation- derivation not expected), Michaelis-Menten constant, enzyme affinity, temperature and pH on reaction velocity of enzyme catalyzed reactions. Enzyme specificity- different types, enzyme activation.	
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Practicals 30 hrs- Essential Experiments (15 hrs) , Group Work (15 hrs)

Essential Experiments

1. Tests for amino acids: Solubility, ninhydrin reaction, xanthoproteic reaction, Millons test, Morners test, glyoxalic acid test, Ehrlich's test, nitroprusside test, lead acetate, test for methionine, aldehyde test, Sakaguchi reaction and isatintes
2. Tests for proteins: Solubility, Ninhydrin reaction, Xanthoproteic reaction, Folin's test, Lowry's test, Biuret test, Heat denaturation, TCA precipitation, Alcohol precipitation.
3. Demonstration of Kinetics of Urease / Trypsin (Effect of pH, substrate Concentration, enzyme concentration• and temperature) .
4. Progress curve of Urease/Trypsin•
5. Digestion of carbohydrates –action of salivary amylase•

Suggested Readings:

1. Lehninger Principles of Biochemistry, 4th Edition by David L. Nelson David L. Nelson (Author)
2. Biochemistry (2004) by Donald Voet, Judith G. Voet Publisher: John Wiley & Sons Inc
3. TextBook of Biochemistry, 5th edition by DM Vasudevan and Sreekumar S, JAYPEE Publishers, New Delhi
4. Experimental Biochemistry: A Student Companion, Beedu Sasidhar Rao & Vijay Deshpande (ed), I.K International Pvt. LTD, New Delhi .
5. Introductory Practical biochemistry, S. K. Sawhney & Randhir Singh (eds) Narosa Publishing House, New Delhi,
6. Standard Methods of Biochemical Analysis, S. K. Thimmaiah (Ed), Kalyani Publishers, Ludhiana.
7. ES West, WR Todd, HS Mason and JT van Bruggen. A text Book of Biochemistry, Oxford and IBH Publishing Co., New Delhi, 1974.
8. E.S. West, W.R. Todd, H.S. Mason and J.T. van Bruggen, A Text Book of Biochemistry, Oxford and IBH Publishing Co., New Delhi, 1974
9. Principles and Techniques of Practical Biochemistry by Keith M. Wilson, John M. Walker Cambridge University Press

No.	Upon completion of the course the graduate will be able to	Cognitive Level	PSO addressed
CO-1	Understanding how chemical constituents of life in a biotechnological perspective	U	PSO-1,2

CO-2	Explain biomolecule modifications and biological significance of various bioconjugates	R, U,Ap	PSO3,4
CO3	Understand complex metabolic pathways in living cells	R , U	PSO3
CO4	Evaluate various tests for presence of amino acids, carbohydrates on the basis of qualitative tests	U, Ap	PSO3,4

R-Remember, U-Understand, Ap-Apply, An-Analyse, E-Evaluate, C-Create

Note: 1 or 2 COs/module

Name of the Course: Metabolism and energetic Credits: 3:0:2 (Lecture:Tutorial:Practical)

CO No.	CO	PO/PSO	Cognitive Level	Knowledge Category	Lecture (L)/Tutorial (T)	Practical (P)
CO-1	Understand the basic constituents of Life	PSO-1,2	U	F, C	L	
CO-2	Explain how biomolecules function and its various structural and functional modifications	PSO3,4	R, U,Ap	P	L	
CO3	Understand the basics of metabolic networks	PSO3	R , U	F	L	
CO4	Evaluate the tests for important biomolecules	PSO3,4	U, Ap	P	L	P

F-Factual, C- Conceptual, P-Procedural, M-Metacognitive

Mapping of COs with PSOs and POs :

	PSO1	PSO2	PSO3	PSO4	PSO5	PSO6	PO1	PO2	PO3	PO4	PO5	PO6
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CO 1	2	2	-	-	-	-						
CO 2	-	-	3	3	-	-						
CO 3	-	-	2	-	-	-						
CO 4	-	-	2	3	-	-						

Correlation Levels:

Level	Correlation
-	Nil
1	Slightly / Low
2	Moderate / Medium
3	Substantial / High

Assessment Rubrics:Continuous Comprehensive Assessment:

Formative : Practical /problem/Survey/Design/Field trip based Assignment in the form of Individual or team work regarding the theory part/Follow any of the tool regarding the above practical assignment-1.Observation of practical skill/Viva/.Quiz/ In class discussions/Good Lab practices/Lab work book

Summative:Follow any of the tool - Written test (theory based)/.Open Book Test(theory based)/ Individual /team work report practical assignmen/Case study report /Literature survey/Individual / Team project report oral presentation/ Laboratory report

End Semester Examination Mapping of COs to Assessment Rubrics :

	Internal Exam	Assignment	Observation of Practical skills	End Semester Examinations
CO 1	✓	✓		✓
CO 2	✓	✓		✓
CO 3	✓	✓		✓
CO 4			✓	✓



University of Kerala

Discipline	BIOTECHNOLOGY				
UK3DSCBIT Code	UK3DSCBIT201				
Course Title	MICROBIOLOGY				
Type of Course	DSC				
Semester	III				
Academic Level	200 – 299				
Course Details	Credit	Lecture per week	Tutorial per week	Practical per week	Total Hours/Week
	4	3 hours	0	2 hours	5
Pre-requisites	Basic knowledge of Biology and Biochemistry				
Course Summary	The Microbiology course delves deeply into the microbial world, covering key aspects from past discoveries to practical applications in various fields. It equips students with a strong foundation in microbiology, necessary lab skills, and a thorough understanding of microbial functions, uses, and control methods vital in scientific and industrial contexts.				

Detailed Syllabus:

Module	Unit	Content	Hrs
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I	Introduction to the microbial world		10
	1	Overview of key events and discoveries in the history of microbiology. Contributions of Anton van Leeuwenhoek, Louis Pasteur and Robert Koch	
	2	Introduction to the prokaryotic world, eukaryotic microorganisms, acellular microorganisms (Viruses, Viroids, Prions, Mycoplasma)	
	3	Introduction to systems of classification: Binomial Nomenclature, Introduction to Bergey's manual	
	4	Nutritional classification of bacteria, Energy production in bacteria (Brief account)	
II	Principles of microbial control		6
	5	General principles: Control by removal, inhibition and sterilisation.	
	6	Physical Methods of Microbial Control : Heat- dry and moist, filtration, radiation	
	7	Chemical Methods of Microbial Control Types and Modes of Action of chemical sterilants and disinfectants	
III	Microbial physiology and culture		12
	9	Ultrastructure of bacteria: Cell wall (Gram positive and negative), internal organisation, spores Staining techniques: Simple staining, negative staining, differential staining (Grams staining)	
	10	Motility in bacteria – structure of flagella, their distribution and patterns of motility, Hanging drop test	
	11	Nutritional requirements in bacteria Culture media: components, types of culture media Pure culture methods (pour plate, spread plate, streaking, culture preservation methods (Lyophilisation, deep freezing)	
	12	Bacterial growth curve, factors affecting the growth of microbes.	
IV	Bacterial Genetics and recombination		10
	13	Bacterial Chromosome, Plasmids- types of plasmids Bacterial mutation and Repair	
	14	Bacterial Recombination- Transformation, Transduction (Generalised and Specialised), Conjugation	
V	Applied Microbiology		7
	17	Agricultural microbiology: Biological nitrogen fixation, Mycorrhizal associations,	
	18	Microbes as biofertilizer – types and application	
	19	Microbes in extreme condition: role of Methanogenic bacteria, extremophiles – Thermophiles, Acidophiles, Halophiles and Alkalophiles, Applications.	
	20	Case study and industrial visit	

Practicals 30hrs-Essential Experiments(15hrs), Group Work(15hrs) Essential

Experiments

1. Handling Microscope
2. Preparation of smear on slide and focusing on microscope (low power and high power objective).
3. Sterilization and aseptic techniques-preparation and sterilization of glassware and solutions, Autoclaving, Hot air oven
4. Media Preparation-Preparation of Luria-Bertani medium, Nutrient agar and their sterilization (Broth and plates).
5. Serial dilution of bacterial cultures and spread plating (L-rod) to find out population density of microbes in a given sample, incubation and observation of colonies
6. Examination of microbial flora of the available soil and water samples. a) Pour plate method, b) Streak plate method - Continuous, Quadrant & T streak.
7. Staining of bacteria-Gram staining, Acid fast staining, Negative staining.
8. Microscopic tests for bacterial motility-Hanging Drop experiment
9. Identification of bacterial and fungal cultures microscopically.
10. Antibiotics sensitivity assays-Kirby Bauer Method

Suggested Readings:

1. A Textbook of Microbiology-P. Chakraborty, New Central Book Agency Pvt. Ltd, Calcutta
2. Modern concept of Microbiology-DD Kumar, SKumar; Vikas Publishing House Pvt. Ltd. New Delhi
3. Introduction to Genetic Engineering & Biotechnology-A.J. Nair; Jones & Bartlett Publishers, Boston, USA.

4. Introduction to Microbiology-J Heritage, EGVEvans, RAKillington; Cambridge University Press.
5. Microbiology–LM Prescott, Brown Publishers, Australia
6. Advances in Microbiology–JPTewari, TNLakhanpal, ISingh, RGupta and BP Chanola; A P H Publishing Corporation, New Delhi.
7. Microbiology: Principles and Explorations–Jacquelyn G. Black. Prentice Hall, New Jersey.

Course Outcomes

No.	Upon completion of the course the graduate will be able to	Cognitive Level	PSO addressed
CO- 1	Understanding the key events in microbial history.	U	PSO-1,2
CO- 2	Explains scope of microbiology and different nutritional types of microorganisms and classification	An	POS1
CO- 3	Understand microbial genetics and metabolism, analyse the recombination methods in bacteria	E	PSO1
CO- 4	Understand strategies for microbial control. Evaluate the role of sterilants and disinfectants.	E	POS1, PSO3
CO- 5	Differentiate between types of culture media and understand factors affecting microbial growth.	C	POS1, PSO4
CO- 6	Gain practical skills in microbial sampling, isolation, staining and culture methods.	C	PSO3

R-Remember, U-Understand, Ap-Apply, An-Analyse, E-Evaluate, C-Create

Note: 1 or 2 COs/module

Name of the Course: MICROBIOLOGY credits: 3:0:2 (lecture:tutorial:practical)

CO No.	CO	PO/PSO	Cognitive Level	Knowledge Category	Lecture (L)/Tutorial (T)	Practical (P)
CO-	Understanding the key events in microbial history.	PSO-1,2	U	F,C	L	-
CO No.	CO	PO/PSO	Cognitive Level	Knowledge Category	Lecture (L)/Tutorial (T)	Practical (P)
CO-2	Explain scope of microbiology and different nutritional types of microorganisms and classification	POS1	An	P	L	-
CO-3	Understand microbial genetics and metabolism, analyse the recombination methods in bacteria	PSO1	E	F	L	-
CO-4	Understand strategies for microbial control. Evaluate the role of sterilants and disinfectants.	POS1,PSO3	E	F	L	
CO-5	Differentiate between types of culture media and understand factors affecting microbial growth.	POS1,PSO4	C	P	L	P

CO-6	Gain practical skills in microbial sampling, isolation, staining and culture methods.	PSO3	C	P	L	P
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F-Factual,C-Conceptual,P-Procedural,M-Metacognitive Mapping of

COs with PSOs and POs :

	PSO1	PSO2	PSO3	PSO4	PSO5	PSO6	PO1	PO2	PO3	PO4	PO5	PO6
CO1	3	-	-	-	-	-						
CO	3	-	-	-	-	-						
	PSO1	PSO2	PSO3	PSO4	PSO5	PSO6	PO1	PO2	PO3	PO4	PO5	PO6
2												
CO3	3	-	-	-	-	-						
CO4	2	-	2	-	-	-						
CO5	2	-	-	3	-	-						
CO6	-	-	3	2	-	-						

CorrelationLevels:

Level	Correlation
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-	Nil
1	Slightly/Low
2	Moderate/Medium
3	Substantial/ High

Assessment Rubrics:

Continuous Comprehensive Assessment:

Formative:

- Interactive Quiz/Group Discussions/Assignment/Student Seminar
- Observation of practical skills/Journal Club presentations
- Punctuality in lab, and time management in completing assigned laboratory tasks

Summative-Internal test papers/Laboratory book/report/Periodic lab tests

Mapping of CO to Assessment Rubrics:

	Internal Exam	Assignment	Observation of Practical skills	End Semester Examinations
CO 1	✓			✓
CO 2	✓	✓		✓
CO 3	✓	✓		✓
CO 4		✓		✓
CO 5		✓		✓
CO 6			✓	✓



University of Kerala

Discipline	BIOTECHNOLOGY				
Course Code	UK3DSCBIT202				
Course Title	BASICS OF ENZYMOLOGY				
Type of Course	DSC				
Semester	III				
Academic Level	200 - 299				
Course Details	Credit	Lecture per week	Tutorial per week	Practical per week	Total Hours/Week
	4	3 hours	0	2 hours	5
Pre-requisites	Basic knowledge of Biology, Biochemistry & Cell biology				
Course Summary	The Basics of Enzymology course provides a comprehensive introduction to the fundamental principles governing enzyme structure, function, kinetics, and regulation. Through a combination of lectures, laboratory sessions, and readings, students will gain a deep understanding of enzymatic mechanisms and their significance in biological processes. The course emphasizes on both theoretical knowledge and practical skills necessary for studying enzymes in various biochemical contexts.				

Detailed Syllabus:

Module	Unit	Content	Hrs
I	Nomenclature, Classification & Purification of Enzymes		8
	1	The Enzyme Commission's system of classification and nomenclature	
	2	Extraction and Purification of Enzymes: Extraction of soluble and membrane bound enzymes; Overview of enzyme purification methods (Precipitation, Electrophoresis, Chromatography), Criteria of enzyme purity	
II	Structure and General properties of enzymes		10
	4	Enzymes and Co-factors, Specificity of enzyme action -Types of specificity, the active site, Lock and key hypothesis, Induced fit hypothesis.	
	5	Mechanism of Catalysis: Acid Base catalysis, Covalent catalysis, Electrostatic catalysis, Enzyme Catalysis	

	6	Factors affecting enzyme activity (substrate concentration, temperature, pH)	
	7	Isozymes, Metalloenzymes, Multienzyme complexes, Synthetic Enzymes.	
III	Kinetics of Enzyme-catalyzed reactions		10
	8	Kinetics of enzyme catalyzed reactions: Factors affecting enzyme kinetics, Significance of Michaelis-Menton constant, Lineweaver–Burk plot	
	9	Kinetics of multi-substrate enzyme - catalyzed reactions – Ping-pong and random order mechanisms	
	10	Enzyme inhibition: Mechanism of enzyme inhibition –Competitive, non –competitive, Uncompetitive and Irreversible Inhibition	
IV	Regulatory mechanisms in enzyme catalysis		8
	11	Allosteric Regulation and Feedback Regulation, Important metabolic pathways regulated by allosteric enzymes (Glycolysis, Krebs Cycle)	
V	Enzyme technology		9
	14	Introduction to Enzyme technology, Enzyme engineering, Computational enzyme design	
	15	Applications of Enzymes: Applications in medicine (diagnostic enzymes, therapeutic enzymes), Applications in Genetic Engineering (Restriction endonucleases, Ligases), Applications in Industry	
	16	Immobilization of enzymes and their applications	

Practical 30 Hours- Essential Experiments (15 hrs) , Group Work (15 hrs)

Essential Experiments

1. Demonstration of Kinetics of Urease / Trypsin (Effect of pH, substrate Concentration, enzyme concentration and temperature).
2. Progress curve of Urease/Trypsin
3. Digestion of carbohydrates –action of salivary amylase

Suggested Readings:

1. "Lehninger Principles of Biochemistry" by David L. Nelson and Michael M. Cox - This comprehensive book covers enzymology along with other fundamental concepts in biochemistry.
2. "Enzymes: Biochemistry, Biotechnology, Clinical Chemistry" by Trevor Palmer - A thorough textbook covering the principles of enzyme structure, function, kinetics, and regulation, as well as applications in biotechnology and clinical chemistry.

3. "Principles of Enzymology for the Food Sciences" by John R. Whitaker, Alphons G.J. Voragen, and Dominic W.S. Wong - Focuses on the enzymology relevant to the food industry, covering topics such as enzyme kinetics, enzyme inhibition, and enzyme applications in food processing.
4. "Introduction to Enzyme and Coenzyme Chemistry" by T. D. H. Bugg - A concise introduction to the basic principles of enzyme and coenzyme chemistry, suitable for students with a background in chemistry or biochemistry.
5. "Enzyme Kinetics: Catalysis and Control" by Daniel L. Purich - Provides an in-depth understanding of enzyme kinetics, including detailed discussions on reaction mechanisms, rate equations, and enzyme inhibition.
6. "Biochemical Calculations: How to Solve Mathematical Problems in General Biochemistry" by Irwin H. Segel - Although not solely focused on enzymology, this book provides valuable guidance on mathematical aspects of enzyme kinetics and other biochemical calculations.
7. "Practical Enzymology" by Hans Bisswanger - Offers practical guidance on experimental techniques in enzymology, including enzyme purification, assay methods, and data analysis.

Course Outcomes

Course Outcomes

No.	Upon completion of the course the graduate will be able to	Cognitive Level	PSO addressed
CO- 1	Understand Enzyme nomenclature, Classification, Extraction and Purification of Enzymes	U, Ap	PSO-1,2
CO- 2	Understand the Structure and General properties of enzymes and enzyme specificity	R, E	PSO1
CO- 3	Explain Kinetics of enzyme catalyzed reactions and factors affecting enzyme catalyzed reactions	U, An	PSO1, PSO3
CO- 4	Understand Regulatory Mechanisms in Enzyme Catalysis and metabolic regulations	E, C	PSO1
CO- 5	Understand enzyme technology and the applications of enzymes in various fields	Ap	PSO3,4

R-Remember, U-Understand, Ap-Apply, An-Analyse, E-Evaluate, C-Create

Note: 1 or 2 COs/module

Name of the Course: Basics of enzymology Credits: 3:0:2 (Lecture:Tutorial:Practical)

CO No.	CO	PO/PSO	Cognitive Level	Knowledge Category	Lecture (L)/Tutorial (T)	Practical (P)
CO- 1	Understand Enzyme nomenclature, Classification, Extraction and	PSO-1,2	U, Ap	F, C	L	
CO- 2	Understand the Structure and General properties of enzymes and enzyme specificity	PSO1	R, E	P	L	
CO- 3	Explain Kinetics of enzyme catalyzed reactions and factors affecting enzyme catalyzed	PSO1, PSO3	U, An	F,P	L	
CO- 4	Understand Regulatory Mechanisms in Enzyme Catalysis and metabolic	PSO1	E, C	F	L	
CO- 5	Understand enzyme technology and the applications of enzymes in various	PSO 3,4	Ap	P	L	P

F-Factual, C- Conceptual, P-Procedural, M-Metacognitive Mapping of COs with PSOs and POs :

	PSO1	PSO2	PSO3	PSO4	PSO5	PSO6	PO1	PO2	PO3	PO4	PO5	PO6
CO 1	2	3	-	-	-	-						
CO 2	3	-	-	-	-	-						
CO 3												
CO 4	3	-	-	-	-	-						

CO 5	-	-	3	2	-	-							
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Correlation Levels:

Level	Correlation
-	Nil
1	Slightly / Low
2	Moderate / Medium
3	Substantial / High

Assessment Rubrics:

Continuous Comprehensive Assessment:

Formative : Practical /problem/Survey/Design/Field trip based Assignment in the form of Individual or team work regarding the theory part/Follow any of the tool regarding the above practical assignment-1.Observation of practical skill/Viva/.Quiz/ In class discussions/Good Lab practices/Lab work book

Summative:Follow any of the tool - Written test (theory based)/.Open Book Test(theory based)/ Individual /team work report practical assignmen/Case study report /Literature survey/Individual / Team project report oral presentation/ Laboratory report

Mapping of COs to Assessment Rubrics :



University of Kerala

Discipline	BIOTECHNOLOGY				
Course Code	UK3DSCBIT203				
Course Title	MICROBIAL METABOLISM				
Type of Course	DSC				
Semester	III				
Academic Level	200–299.				
Course Details	Credit	Lecture per week	Tutorial per week	Practical per week	Total Hours/Week
	4	3 hours	0	2 hours	5
Pre-requisites	Basic knowledge in Microbiology and Biochemistry				
Course Summary	Microbial metabolism is a fundamental aspect of microbiology that explores the biochemical pathways and mechanisms by which microorganisms obtain energy, grow and interact with their environments. This graduate-level course delves into the intricate world of microbial metabolic processes, emphasizing the diversity of metabolic strategies employed by bacteria, archaea, fungi, and other microorganisms.				

Detailed Syllabus:

Module	Unit	Content	Hrs
I	Nutritional classification & Nutrient transport in microbes		6
	1	Nutritional classification of bacteria	
	2	Nutrient transport across the cell: Diffusion: Passive and facilitated; Primary active and secondary active transport	
	3	Group translocation (phosphotransferase system) electroneutral transport; transport of Iron.	
II	Photosynthesis & Respiration in Bacteria		10
	4	Photosynthetic pigments of bacteria- chlorophyll a and bacteriochlorophyll, carotenoids, phycobiliproteins, leghaemoglobin	
	5	Oxygenic and anoxygenic photosynthesis Mechanism of photosynthesis in bacteria (purple non sulphur bacteria, green sulphur bacteria) and cyanobacteria	
	6	Chemolithotrophy -- oxidation of sulphur, iron, hydrogen & nitrogen Methanogenesis, Bioluminescence	
	7	Respiration in bacteria- aerobic respiration	

		Glycolysis and tricarboxylic acid cycle Electron transport and oxidative phosphorylation in Bacteria Anaerobic respiration- Fermentation- lactic acid and alcohol fermentation, mixed acid fermentation, Lactate fermentation (homofermentative and heterofermentative pathways).	
	8	Assimilation of nitrogen, sulphur, phosphorus	
III	Synthesis of biopolymers		10
	9	Biosynthesis of peptidoglycan, biopolymers, PHB	
	10	Biosynthesis of vitamins, amino acids and nucleotides	
	11	Regulation of metabolic pathways	
	12	Overview of Microbial metabolites-marine sources	
IV	Biochemical characterization of bacteria		10
	13	Importance of Biochemical characterisation Types: Carbohydrate fermentation test, Methyl red test, Citric acid utilization test. (D) Hydrogen sulfide production test.	
	14	Principle of Sugar utilization test, Sugar fermentation test, IMViC test	
	15	Enzyme detection – Catalase, Oxidase, Oxidative-fermentative test	
	16	Gelatinase assay	
V	Industrial importance of Microbial metabolism		9
	17	Microorganisms of industrial importance. Biology of industrial microorganisms: Isolation, Screening and Preservation.	
	18	Fermentation process, Types of fermentation and Downstream processing- recovery and purification of end products of metabolism-a basic account	
	19	Strain improvement of microbes for industrial purposes	
	20	Examples of commercial products of microbial origin- case study	

Practicals 30 hrs- Essential Experiments (15 hrs) , Group Work (15 hrs)

Essential Experiments

1. Effect of temperature, pH, salt, Carbon source and Nitrogen source on the growth of bacteria
2. Study and plot growth curve of E.coli by turbidometric method
3. Demonstration of production of acid and gas during lactose fermentation
4. Urease test
5. Gelatin hydrolysis
6. Isolation and culture of photosynthetic bacteria.
7. Starch hydrolysis test by amylase producing microbes, and enzyme assay.

Suggested Readings:

1. Madigan MT, and Martinko JM (2014). Brock Biology of Microorganisms. 14th edition. Prentice Hall International Inc.

2. Moat AG and Foster JW. (2002). Microbial Physiology. 4th edition. John Wiley & Sons
3. Nelson David L and Cox Michael M, Lehninger, Principles of Biochemistry, Macmillan Press, Worth Publishers, New Delhi.
4. Reddy SR and Reddy SM. (2005). Microbial Physiology. Scientific Publishers India
5. Sambrook J and Russell DW. (2001). Molecular Cloning: A Laboratory Manual. 4th Edition, Cold Spring Harbour Laboratory press.
6. Stanier RY, Ingrahm JI, Wheelis ML and Painter PR. (1987). General Microbiology. 5th edition, MacMillan Press.
7. Willey JM, Sherwood LM, and Woolverton CJ. (2013). Prescott's Microbiology. 9th edition. McGraw Hill Higher Education.

Course Outcomes

No.	Upon completion of the course the graduate will be able to	Cognitive Level	PSO addressed
CO-1	Understanding of bacterial nutritional requirements to classify bacteria & predict their growth conditions.	U, R	PSO-1,2
CO-2	Describe different mechanisms of nutrient transport across cell membranes & evaluate their significance in bacterial metabolism.	U, E	PSO1,3
CO-3	To know the process of photosynthesis in bacteria	U, R	PSO1
CO-4	Examine the environmental significance of chemolithotrophy	U, An	PSO3
CO-5	Compare the efficiency of aerobic & anaerobic respiration pathways in bacteria by comparing their energy yields.	U	PSO1
CO-6	Explain the regulatory mechanisms involved in biosynthetic pathways	U	PSO3,4
CO-7	Apply their practical skills to identify bacteria using different biochemical tests	Ap, An	PSO5

R-Remember, U-Understand, Ap-Apply, An-Analyse, E-Evaluate, C-Create

CO 1	2	2	-	-	-	-						
CO 2	2	-	3	-	-	-						
CO 3	2	-	-	-	-	-						
CO 4	-	-	2	-	2	-						
CO 5	2	-	-	-	-	-						
CO 6	-	-	3	3	-	-						
CO7					1							

Correlation Levels:

Level	Correlation
-	Nil
1	Slightly / Low
2	Moderate / Medium
3	Substantial / High

Assessment Rubrics:Continuous Comprehensive Assessment: Formative :

Formative : Practical /problem/Survey/Design/Field trip based Assignment in the form of Individual or team work regarding the theory part/Follow any of the tool regarding the above practical assignment-1.Observation of practical skill/Viva/.Quiz/ In class discussions/Good Lab practices/Lab work book

Summative:Follow any of the tool - Written test (theory based)/.Open Book Test(theory based)/ Individual /team work report practical assignmen/Case study report /Literature survey/Individual / Team project report oral presentation/ Laboratory report

Mapping of COs to Assessment Rubrics :

	Internal Exam	Assignment	Observation of Practical skills	End Semester Examinations
CO 1	✓	✓	23	✓
CO 2	✓			✓

CO 3	✓			✓
CO 4	✓	✓		✓
CO 5	✓	✓		✓
CO 6	✓			✓
CO7			✓	✓



University of Kerala

Discipline	BIOTECHNOLOGY				
Course Code	UK3DSCBIT204				
Course Title	PLANT PHYSIOLOGY				
Type of Course	DSC				
Semester	III				
Academic Level	200 -299.				
Course Details	Credit	Lecture per week	Tutorial per week	Practical per week	Total Hours/Week
	4	3 hours	0	2	5
Pre-requisites	Basic knowledge on life science, specifically on physiological processes in plant body. Fundamentals of Biology, Biochemistry				
Course Summary	This course gives a fundamental knowledge on the biophysical and biochemical processes that function in a plant system. The students will learn the basic mechanisms governing the life processes of plants at a cellular, molecular, and whole-organism level.				

Detailed Syllabus:

Module	Unit	Content	Hrs
I	Introduction to plant physiology		8
	1	Physiological process, their significance and other application: Water relations and Mineral nutrition in plant body	
11	Photosynthesis		12
	2	Photosynthetic apparatus and pigment systems-chromatographic techniques for the separation of photosynthetic pigments, Raw materials of photosynthesis Light perception and signal transduction	
	3	a) Light reaction i) Radiant energy and its effects on chlorophyll pigments ii) Cyclic and non-cyclic photophosphorylation iii) Source of oxygen liberated iv) Hill reaction	
	4	b) Dark reaction i) Trace the path of carbon in photosynthesis ii) Calvin cycle iii) C3 and C4 plants. CAM plants. iv) Photorespiration v) Factors affecting photosynthesis. Law of limiting factors	

	5	Definition and general equation. Significance, Respiratory substrates, Mechanism - Glycolysis, Krebs cycle, terminal oxidation. Oxidative pentose phosphate pathway, Factors affecting respiration, Anaerobic respiration	
III	Growth and Development		10
	6	Seed germination and seedling establishment -Dormancy and germination of seeds. Flowering and reproductive development, Differentiation, morphogenesis Hormonal regulation of plant growth - Auxins, Gibberellins, Cytokinins, Abscissic acid, Ethylene and their practical applications	
	7	Senescence and programmed cell death	
	8	Signal Transduction and Plant Responses: how plants perceive and respond to internal and external signals. receptor proteins, intracellular signaling pathways, gene expression regulation, and the integration of multiple signaling inputs.	
IV	Plant-biotic- abiotic Interactions		6
	9	Interactions between plants and microorganisms, beneficial symbioses (e.g., mycorrhizae, nitrogen-fixing bacteria) and pathogenic infections.	
	10	Biotic stress responses (pathogens, herbivores)- Molecular communication, defense mechanisms, and the implications for plant health and agriculture	
	11	Abiotic stress responses (drought, salinity, temperature) Circadian rhythms and biological clocks	
V	Biotechnological Applications		9
	12	Crop improvement through biotechnology, Biofortification and nutritional enhancement	
	12	Phytoremediation and bioremediation	

Practicals 30 hrs- Essential Experiments (15 hrs) , Group Work (15 hrs)

Essential Experiments

1. Evolution of oxygen during photosynthesis
2. Necessity of chlorophyll, CO₂ and light in photosynthesis
3. Measurement of photosynthesis.

4. Simple respiroscope , Respirometer of R.Q. , Anaerobic respiration
5. Geotropism and phototropism Klinostät ,. Hydrotropism
6. Measurement of growth Arc or Lever Auxonometer
- 7.Plant tissue culture and transformation methods
8. Bioinformatics analysis of plant genomic data

Suggested Reading:

1. Devlin & Witham Plant Physiology, C B S publishers.
2. Devlin R.M. (1979) Plant Physiology.
3. Dieter Hess (1975): Plant physiology.
4. Jain V. K. (1996) Fundamentals of Plant Physiology.
5. Kochhar P. L. & Krishnamoorthy H. N. Plant Physiology. Atmaram & Sons Delhi, Lucknow.
6. Kumar & Purohit Plant Physiology - Fundamentals and Applications, Agrobotanical publishers.
7. Malik C. P. & Srivastava A. K. Text book of Plant Physiology Kalyani Publishers New Delhi.
8. Noggle G R & Fritz G J (1991) Introductory Plant physiology, Prentice Hall of India
9. Pandey S.N. & Sinha B. K. (1986) Plant physiology, Vikas publishing House- New Delhi.
10. Salisbury F.B and Ross C.W. (2006): Plant Physiology 4Edn, Wadsworth publishing company.
11. Sundara Rajan S. College Botany Vol. IV, Himalaya publishing House.

Course Outcomes

	Upon completion of the course the graduate will be able to	Cognitive Level	PSO addressed
CO-1	Understand the physiological process of plant physiological process and molecular signalling involved	R, U	PSO1
CO-2	Aware the concept of photosynthesis	R, U	PSO3
CO-3	Analyse the significance of respiration	U,An	PSO1,3

CO-4	Understand the signal transduction pathways and hormonal regulation involved in growth and development	U,Ap	PSO3,4
CO-5	Understand the application of physiological process	U,An	PSO3

R-Remember, U-Understand, Ap-Apply, An-Analyse, E-Evaluate, C-Create

Note: 1 or 2 COs/module

Name of the Course: Plant physiology Credits: 3:0:2 (Lecture:Tutorial:Practical)

CO No.	CO	PO/PSO	Cognitive Level	Knowledge Category	Lecture (L)/Tutorial (T)	Practical (P)
CO-1	Understand the physiological process of plant physiological process and molecular signalling involved	PSO1	R, U	F, C	L	
CO-2	Aware the concept of photosynthesis	PSO3	R, U	P	L	
CO-3	Analyse the significance of respiration	PSO1,3	U,An	C	L	P
CO-4	Understand the signal transduction pathways and hormonal regulation involved in growth and development	PSO-3,4	U,Ap	C	L	
CO-5	Understand the application of physiological process	PSO3	U,An	P	L	

F-Factual, C- Conceptual, P-Procedural, M-Metacognitive

Mapping of COs with PSOs and POs :

	PSO1	PSO2	PSO3	PSO4	PSO5	PSO6	PO1	PO2	PO3	PO4	PO5	PO6
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CO 1	2	-	-	-	-	-						
CO 2	-	-	3	-	-	-						
CO 3	2	-	3	-	-	-						
CO 4	-	-	2	2	-	-						
CO 5	-	-	3	-	-	-						

Correlation Levels:

Level	Correlation
-	Nil
1	Slightly / Low
2	Moderate / Medium
3	Substantial / High

Assessment Rubrics: **Continuous Comprehensive Assessment:**

Formative : Practical /problem/Survey/Design/Field trip based Assignment in the form of Individual or team work regarding the theory part/Follow any of the tool regarding the above practical assignment-1.Observation of practical skill/Viva/.Quiz/ In class discussions/Good Lab practices/Lab work book

Summative:Follow any of the tool - Written test (theory based)/.Open Book Test(theory based)/ Individual /team work report practical assignmen/Case study report /Literature survey/Individual / Team project report oral presentation/ Laboratory reportMapping of COs to Assessment Rubrics :

	Internal Exam	Assignment	Observation of Practical skills	End Semester Examinations
CO 1	✓			✓
CO 2	✓			✓
CO 3	✓			✓
CO 4		✓		✓
CO 5		✓	29	✓



University of Kerala

Discipline	BIOTECHNOLOGY				
Course Code	UK3DSCBIT205				
Course Title	ANIMAL PHYSIOLOGY				
Type of Course	DSC				
Semester	III				
Academic Level	200 - 299.				
Course Details	Credit	Lecture per week	Tutorial per week	Practical per week	Total Hours/Week
	4	3 hours	0	2 hours	5
Pre-requisites	Fundamental Biology, Basic Cell biology				
Course Summary	This course provides an in-depth exploration of the fundamental principles governing the function of the human body, and will answer how our body functions. It examines the intricate mechanisms underlying various physiological processes, ranging from cellular functions to integration of organ systems. Through a combination of lectures, and discussions, students delve into the complex interplay of molecular, cellular, and systemic processes that maintain homeostasis and regulate bodily functions.				

Detailed Syllabus:

Module	Unit	Content	Hrs
I	Nutrition and Respiration		6
	1	Nutrition: Types of nutrition; Mechanical and chemical digestion of carbohydrates, proteins and fats; hormonal control of digestion; absorption mechanism; BMR Vitamin deficiency diseases.	
	2	Respiration: respiratory pigments and their role; gas transport – oxygen and CO ₂ transport; Oxyhaemoglobin curve; Bohr effect; Carbon monoxide poisoning. Physiological effects of smoking	
II	Circulatory system		10
	3	Circulation: Body fluids – importance and types; closed and open types of circulatory system; blood– composition and functions; blood groups – ABO and Rh systems, MN, Lewis and Bombay groups; blood clotting – intrinsic and extrinsic mechanisms and their factors; anticoagulants.	
	4	Heart: Detailed structure and types of heart – tubular and chambered; neurogenic and myogenic; pacemakers and conducting system of human heart; cardiac rhythm; blood pressure; electrocardiogram. Common cardiovascular diseases (hypertension, arteriosclerosis, myocardial infarction), Adaptations for different metabolic demands	
III	Physiology of excretion and muscle movement		10

	5	Excretion: nitrogenous wastes; ammonotelic, ureotelic and uricotelic modes of excretion; structure of human nephron; urine formation in man – detailed account with countercurrent system; normal and abnormal constituents of urine; hormonal regulation of renal function; Dialysis and artificial kidney	
	6	Muscle Physiology: types of muscles; ultrastructure of striated muscle fibre; muscle contraction, Skeletal adaptations for locomotion and support, Biomechanics of movement; chemistry of contraction; neuromuscular junction; fatigue; muscle twitch; latent and refractory periods; rigor mortis	
IV	Nerve Physiology		10
	7	Nerve Physiology: Sense organs-eyes, (physiology of vision), ear (structure and functions- hearing and balancing), olfactory organs and taste receptors;	
	8	Structure of a typical neuron; types of neurons; myelinated and nonmyelinated nerve fibres; structure and types of synapse; initiation and conduction of nerve impulse;	
	9	EEG; Nervous disorders - epilepsy, Alzheimer's disease, Parkinson's disease.	
		Action potentials and neurotransmission- neurotransmitters; synaptic transmission; reflex action and reflex arc Hormonal regulation and signaling pathways, Integration of neural and endocrine control systems	
V	Endocrinology		9
	10	Endocrinology: hormones – definition and types of hormones; mechanism of hormone action-at the levels of cell membrane, organelles and genes; positive and negative feedback regulation	
	11	Structure and functions of endocrine glands – thyroid, parathyroid, thymus, islets of Langerhans, adrenal, pituitary, hypothalamus, pineal body, gonads and placenta; brief account of prostaglandins	
	12	Hormonal disorders	

Practicals 30 hrs- Essential Experiments (15 hrs) , Group Work (15 hrs)

Essential Experiments

1. Paper partition chromatography of amino acids (3 amino acids and a mixture)
2. Blood smear preparation – identification of leukocytes
3. Determination of human blood group – A, B, AB and O, and Rh+ and Rh-
4. Osmotic properties of RBCs – effect of isotonic, hypotonic and hypertonic solutions.
5. Activity of human salivary amylase on starch
6. Detection of Abnormal constituents of urine (glucose and albumin).
7. Estimate the butterfat content of raw milk, collected fresh from different animal sources
8. Working in groups, chart out physiological adaptations in animals
9. Subject students groups to animal physiology simulations illustrating hierarchies in organisation
10. Separation of serum proteins by gel filtration.

Suggested Readings

- 1.Arora, Mohan P. Animal Physiology. Himalaya Publishing House
- 2.Mariakuttikan and Arumugam, N. Animal Physiology. Saras Publication
- 3.Nagabhushanam, R. et al. Textbook of Animal Physiology. Oxford & IBHS
- 4.Rastogi, S.C. Essentials of Animal Physiology. Wiley Eastern Ltd.
- 5.Sebastian, M.M. Animal Physiology. Madonna Books, Kottayam
- 6.Verma, P.S. Tyagi, B.S. and Agarwal, V.K. Animal Physiology. S.Chand & Co.
- 7.Berry, A.K. A Text book of Animal Physiology, Emkay Publications.
- 8.Best and Taylor's Physiological Basis of Medical Practice. West, J.B. (Ed.) B.I. Waverly.

Course Outcomes

No.	Upon completion of the course the graduate will be able to	Cognitive Level	PSO addressed
CO-1	Understand importance of nutrition and associated disorders	U	PSO-1,2
CO-2	Understand physiology of important vital organs and mechanism of function various parts	R, U	PSO1
CO3	Understand various physiological disorders associated with important organs and diagnosis	R, U	PSO3,4
CO4	Describe the effect of hormone action- at cellular level and structure and functions of endocrine glands	R, U	PSO3

R-Remember, U-Understand, Ap-Apply, An-Analyse, E-Evaluate, C-Create

Note: 1 or 2 COs/module

Name of the Course: Animal physiology

Credits: 3:0:2 (Lecture:Tutorial:Practical)

CO No.	CO	PO/PSO	Cognitive Level	Knowledge Category	Lecture (L)/Tutorial (T)	Practical (P)
CO-1	Understand importance of nutrition physiological functions	PSO-1,2	U	F, C	L	

CO-2	Understand animal physiology	PSO1	R, U	P	L	
CO3	Understand various physiological disorders	PSO3,4	R, U	F	L	
CO4	Understand How hormones controls the physiological functions	PSO3	R, U	C	L	

F-Factual, C- Conceptual, P-Procedural, M-Metacognitive

Mapping of COs with PSOs and POs :

	PSO1	PSO2	PSO3	PSO4	PSO5	PSO6	PO1	PO2	PO3	PO4	PO5	PO6
CO 1	1	3	-	-	-	-						
CO 2	3	-	-	-	-	-						
CO 3	-	-	2	2	-	-						
CO 4	-	-	2	-	-	-						

Correlation Levels:

Level	Correlation
-	Nil
1	Slightly / Low
2	Moderate / Medium
3	Substantial / High

Assessment Rubrics:Continuous Continuous Comprehensive Assessment:

Formative : Practical /problem/Survey/Design/Field trip based Assignment in the form of Individual or team work regarding the theory part/Follow any of the tool regarding the above practical assignment-1.Observation of practical skill/Viva/.Quiz/ In class discussions/Good Lab practices/Lab work book

Summative:Follow any of the tool - Written test (theory based)/.Open Book Test(theory based)/ Individual /team work report practical assignmen/Case study report /Literature survey/Individual / Team project report oral presentation/ Laboratory report

	Internal Exam	Assignment	Observation of Practical skills	End Semester Examinations
CO 1	✓			✓
CO 2	✓			✓
CO 3	✓	✓		✓
CO 4		✓		✓



University of Kerala

Discipline	BIOTECHNOLOGY				
Course Code	UK3DSCBIT206				
Course Title	CELL & MOLECULAR BIOLOGY				
Type of Course	DSC				
Semester	III				
Academic Level	200 - 299				
Course Details	Credit	Lecture per week	Tutorial per week	Practical per week	Total Hours/Week
	4	3 hours	0	2 hours	5
Pre-requisites	Basic understanding of cell structure and function, genetics and regulation				
Course Summary	At the graduate level, a course in Cell Biology and Genetics typically delves deep into the intricate mechanisms underlying cellular function, molecular genetics, and their profound implications in various biological processes				

Detailed Syllabus:

Module	Unit	Content	Hrs
I	Cell organelles and functions		12
	1	History and scope of Cell Biology	
	2	Structure and functions of cell organelles (A brief account only) – Nucleus, Mitochondria, Chloroplast, Ribosomes, Endoplasmic Reticulum, Golgi Bodies, Lysosomes, Centrioles	
	3	Structure, function and components of cell membranes, Fluid mosaic model (A brief account)	
	4	Introduction to cell signalling pathways (Basics of G protein coupled receptors, enzyme linked receptors, ion transporters)	
II	Cell division		4
	5	Mitosis and meiosis	
	6	Cell cycle phases and checkpoints (G1, G2/M, spindle checkpoint), importance of cyclins and cyclin dependant kinases	
III	Replication		8
	7	Organization of DNA in chromosomes, Nucleosome model, Organisation of prokaryotic and eukaryotic gene	
	8	Mechanism of Replication of DNA (prokaryotic and eukaryotic),	

		Enzymes involved in DNA replication	
IV	Gene expression		12
	9	Genetic code, codon, Transcription(Prokaryotic and eukaryotic)–mechanism, Post-transcriptional modification of mRNA (importance of 5' cap and 3'tail), Types of RNA-mRNA, tRNA, rRNA and snRNA, mi RNA.	
	10	Splicing mechanism, self-splicing RNA (Group II Introns)	
	11	Translation- mechanism of translation in prokaryotes and eukaryotes.	
	12	Operon concept (lac and trp operon), catabolic repression, attenuation, control of gene expression and post translational modifications (basics only)	
V	Genetics and Case Studies		9
	13	Mendelian genetics and basics of gene interactions Case study- Classic experiments demonstrating DNA as the genetic material Case Study: Tay-Sachs Disease -Lysosomal Storage Inclusion Case Study - protein misfolding Disease- Misfolded Amyloid-β Protein Case Study-colour variation in pepper, Case study- lethal alleles multiple alleles-ABO Blood group. Case study- PAX6 transcription factor Mutation – Eye Development Disorder (Aniridia))	

Practicals 30 hrs- Essential Experiments (15 hrs), Group Work (15 hrs)

Essential Experiments

1. Study of different types of cells (prokaryotes and eukaryotes) using slides/models/charts.
2. Study of cytoplasmic organelles and cell inclusions (through permanent slides, models and charts)
3. Study of stages of mitosis – squash preparation of onion root tip.
4. Preparation of solutions and buffers for DNA isolation
5. Isolation of Genomic DNA from a suitable source- bacteria, plant or animal tissue
6. Examination of the purity of DNA by agarose gel electrophoresis.
7. Quantification of DNA by UV-spectrophotometer
8. Isolation and purification of plasmid DNA
9. Agarose gel analysis of plasmid DNA

Suggested Readings

1. Powar, C.B. Cell Biology. Himalaya Publishing House.
2. Verma, P.S. & Agarwal, V.K. Cytology, S. Chand & Co.
3. Alberts, B. et al. Molecular Biology of the Cell. Garland Science.
4. DeRobertis, E.D.P. and DeRobertis, E.M.P. Cell and Molecular Biology, Lippincott Williams and Wilkins
5. Karp, Gerald. Cell and Molecular Biology. John Wiley and Sons

6. Lodish,Harveyetal.MolecularCellBiology.ScientificAmericanBooks
7. Burns, G. W. &Bottino, P. J. The Science of Genetics. Maxwell McMillan
8. Curt Stein. Principles of Human Genetics. Euresia Publishing House
9. Gardner, E. J. et al. Principles of Genetics. John Wiley & Sons.
10. Goodenough, U. Genetics. Halt, Reinharts& Winston
11. Gupta, P.K. Cytogenetics. Rastogi & Co.
12. Sinnott, W.E., Dunn, L.C. and Dobzhansky, T. Principles of Genetics, TMH
13. Verma, P.S. and Agarwal V.K. Genetics. S.Chand and Co.
14. Applied Molecular Genetics – R L Miesfeld; Wiley.Liss, New Delhi.
15. Essential molecular Biology- A practical Approach, T. A. Brown; Oxford, New York
16. Gene VIII- Benjamin Lewin; Oxford University Press.

Course Outcomes

No.	Upon completion of the course the graduate will be able to	Cognitive Level	PSO addressed
CO-1	Analyse the cooperationamong cellular organelles ensures proper cellular function , including metabolism, communication, division, and survival	An	PSO-1,2
CO-2	Evaluate the role of cell cycle and its checkpoints controlling optimum cellular functions and Consequences of Checkpoint Failure	E	PSO1
CO-3	Analyse therole of gene interactions affecting phenotypes and Decode the various Chromosomal Anomaliesin human genetic disorders	C, E	PSO1
CO-4	Understand the concept of gene, its structure and expression	Ap	PSO1,PSO3
CO-5	Understand the gene regulation mechanisms in a living cell	An	PSO3,4

R-Remember, U-Understand, Ap-Apply, An-Analyse, E-Evaluate, C-Create

Note: 1 or 2 COs/module

Name of the Course: Cell biology & Genetics Credits: 3:0:2 (Lecture:Tutorial:Practical)

CO No.	CO	PO/PSO	Cognitive Level	Knowledge Category	Lecture (L)/Tutorial (T)	Practical (P)
			37			

CO-1	Analyse the cooperation among cellular organelles ensures proper cellular function , including metabolism, communication, division, and survival	An	An	F, C	L	
CO-2	Evaluate the role of cell cycle and its checkpoints controlling optimum cellular functions and Consequences of Checkpoint Failure		E	F C	L	P
CO-3	Analyse the role of gene interactions affecting phenotypes and Decode the various Chromosomal Anomalies in human genetic disorders	PSO1	C	P	L	
CO-4	Understand the concept of gene, its structure and expression	PSO1	Ap	C	L	
CO-5	Understand the gene regulation mechanisms in a living cell	PSO1, PSO3	An		L	

F-Factual, C- Conceptual, P-Procedural, M-Metacognitive

Mapping of COs with PSOs and POs :

	PS O1	PSO2	PSO3	PSO4	PSO5	PSO6	PO1	PO2	PO3	PO4	PO5	PO6
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CO 1	2	3	-	-	-	-						
CO 2	2	-	-	-	-	-						
CO 3	2	-	-	-	-	-						
CO 4	2	-	2	-	-	-						
CO 5	-	-	2	3	-	-						

Correlation Levels:

Level	Correlation
-	Nil
1	Slightly / Low
2	Moderate / Medium
3	Substantial / High

Assessment Rubrics: Continuous Comprehensive Assessment: Formative :

- Interactive Quiz ,Group Discussions,Assignment,Student Seminar
- Observation of practical skills,Journal Club presentations
- Punctuality in lab, and time management in completing assigned laboratory tasks

Summative Internal test papers,Laboratory book/ report,Periodical lab tests

Mapping of COs to Assessment Rubrics :

	Internal Exam	Assignment	Project Evaluation	End Semester Examinations
CO 1	✓			✓
CO 2	✓			✓
CO 3	✓			✓
CO 4	✓	✓		✓
CO 5	✓	✓		✓

University of Kerala

Discipline	BIOTECHNOLOGY				
Course Code	UK3DSCBIT207				
Course Title	MOLECULAR BIOLOGY				
Type of Course	DSC				
Semester	III				
Academic Level	200 - 299.				
Course Details	Credit	Lecture per week	Tutorial per week	Practical per week	Total Hours/Week
	4	3 hours	0	2 hours	5
Pre-requisites	Essentials of Biotechnology Basic understanding on Genetics and DNA structure				
Course Summary	This core-course imparts an essential foundation for understanding of mechanisms and regulations of gene expression at molecular level. Understanding the molecular basis of life is very important to apply manipulation strategies in the future for genetic engineering and genome editing.				

Detailed Syllabus:

Module	Unit	Content	Hrs
I	Molecular basis of life		8
	1	History and significant discoveries in molecular biology, Classic experiments demonstrating DNA as the genetic material	
	2	Central dogma of molecular biology Eukaryotic chromosomes – molecular organization, nucleosomes, telomeres.	
	3	Structure of DNA and types of DNA.	
	4	Replication of DNA (prokaryotic and eukaryotic), enzymes involved in DNA replication.	
II	Genes, Genetic code and Cytoplasmic Genome		8
	5	Organisation of prokaryotic and eukaryotic gene- split genes, introns and exons, reading frame, enhancers and silencers	
	6	Genetic code - properties of genetic code, Codons, codon assignment, redundancy and wobble concept	
	7	Molecular organization of transposons in prokaryotes and eukaryotes	
	8	Mitochondrial DNA and chloroplast DNA	

III	Gene expression		10
	9	Transcription(Prokaryotic and Eukaryotic)- mechanism. RNA Polymerase, promoter, transcription factors.	
	10	Types of RNA-mRNA, tRNA, rRNA and small nuclear RNA (snRNA), mi RNA.	
	11	Post-transcriptional modification of mRNA in eukaryotes-capping, tailing and splicing mechanisms.	
	12	Translation- mechanism of translation in prokaryotes and eukaryotes	
	13	Post translational modification of proteins	
IV	Gene regulation		10
	13	Prokaryotic gene regulation, operon, (lac and trp operon), catabolic repression, attenuation and anti-termination	
	14	Eukaryotic gene regulation; levels of control of gene expression	
	15	mRNA degradation and protein degradation control,	
	16	RNA interference, microRNAs and siRNA	
	17	Translational regulation	
V	Tools and Techniques in Molecular Biology		9
	18	DNA isolation: Principle and Protocol, Purification and quantification Beer-Lambert law, Colorimeter, Spectrophotometer	
	19	Electrophoresis: Principle of electrophoresis, Native Gel Electrophoresis, SDS-PAGE, Agarose gel electrophoresis.	
	20	PCR, Blotting techniques, Microarray, Flow cytometry	
	21	CRISPR-Cas9 system: mechanism and applications	

Practicals 30 hrs- (Essential Experiments (15 hrs), Group Work (15 hrs))

Essential Experiments

1. Familiarisation of instruments and equipment's used in molecular biology laboratory
2. Preparation of solutions and buffers for DNA isolation
3. Isolation of Genomic DNA from a suitable source- bacteria, plant or animal tissue
4. Examination of the purity of DNA by agarose gel electrophoresis.
5. Quantification of DNA by spectrophotometer
6. Visit a molecular biology laboratory within the entire course tenure

Suggested Reading:

1. Applied Molecular genetics – R L Miesfeld; Wiley.Liss, New Delhi.
2. Essential molecular Biology- A practical Approach, T A Brown; Oxford, New York
3. Gene VIII- Benjamin Lewin; Oxford University Press.
4. Molecular Biology, PS Verma and VK Agarwal, S.Chand& Company pvt Ltd, New Delhi
5. Introduction to Molecular biology- P. Paoella; McGraw Hill, New York

6. Molecular Biology of the gene – Watson, Baker, Bell Gann, Lewinw, Losick; Pearson Education Pvt.Ltd, New Delhi
7. Molecular cell biology, H S Bhamrah; Anmol Publications Pvt. Ltd., New Delhi.
8. PCR 3 - Practical Approach – C. Simon Hearington& John J O’Leary; Oxford, New York
9. Principles of Gene manipulation- R.W.Old& S.B. Primrose; Blackwell Scientific Publications
10. M. R. Green, J. Sambrook. Molecular Cloning: A Laboratory Manual (Cold Spring Harbor, ed. 4, 2012).
11. M. M. Burell. Enzymes of Molecular Biology (Humana Press, 1993).

Course Outcomes

No.	Upon completion of the course the graduate will be able to	Cognitive Level	PSO addressed
CO-1	Understand the concept of gene and its structure	R, U	PSO-1,2
CO-2	Analyse the genome organization up to chromosome level	U, An	PSO-1
CO-3	Illustrate how genetic code determines the amino acid sequence of proteins.	R, U, Ap	PSO1, PSO3
CO-4	Differentiate the mechanism of DNA replication in prokaryotes with eukaryotes	U, An	PSO-2,3
CO-5	Compare & contrast Prokaryotic and Eukaryotic Transcription and translation mechanism	U, E	PSO-3,4
CO-6	Generalise the various gene regulation mechanisms in both prokaryotes and eukaryotes	E, C	PSO-4

R-Remember, U-Understand, Ap-Apply, An-Analyse, E-Evaluate, C-Create

Note: 1 or 2 COs/module

Name of the Course: Molecular Biology Credits: 3:0:2 (Lecture:Tutorial:Practical)

CO No.	CO	PO/PSO	Cognitive Level	Knowledge Category	Lecture (L)/Tutorial (T)	Practical (P)
CO-1	Understand the concept of	PSO-1,2	R, U	F, C	L	

	gene					
CO-2	Analyse the genome organization up to chromosome level	PSO1	U, An	C, P	L	
CO-3	Illustrate how genetic code determines the amino acid sequence of proteins.	PSO1, PSO3	R, U, Ap	F, C	L	
CO-4	Differentiate the mechanism of DNA replication in prokaryotes with eukaryotes.	PSO-2,3	U, An	P		P
CO-5	Compare & contrast Prokaryotic and Eukaryotic Transcription and translation mechanism	PSO-3,4	E	F, C	L, T	
CO-6	Generalise the various gene regulation mechanisms in both prokaryotes and eukaryotes	PSO-4	E, C	C, P		P

F-Factual, C- Conceptual, P-Procedural, M-Metacognitive

Mapping of COs with PSOs and POs :

	PSO1	PSO2	PSO3	PSO4	PSO5	PSO6	PO1	PO2	PO3	PO4	PO5	PO6
CO 1	1	3	-	-	-	-						
CO 2	2	-	-	-	-	-						
CO 3	2	-	3	-	-	-						
CO 4	-	-	2	3	-	-						
CO 5	-	-	3	3	-	-						
CO 6	-	-	3	-	-	-						

Correlation Levels:

Level	Correlation
-	Nil
1	Slightly / Low
2	Moderate / Medium
3	Substantial / High

Assessment Rubrics:

Continuous Comprehensive Assessment:

Formative : Practical /problem/Survey/Design/Field trip based Assignment in the form of Individual or team work regarding the theory part/Follow any of the tool regarding the above practical assignment-1.Observation of practical skill/Viva/.Quiz/ In class discussions/Good Lab practices/Lab work book

Summative:Follow any of the tool - Written test (theory based)/.Open Book Test(theory based)/ Individual /team work report practical assignment/Case study report /Literature survey/Individual / Team project report oral presentation/ Laboratory report

Mapping of COs to Assessment Rubrics :

	Internal Exam	Assignment	Observation of practical skills	End Semester Examinations
CO 1	✓			✓
CO 2	✓			✓
CO 3	✓			✓
CO 4	✓		✓	✓
CO 5		✓		✓
CO 6			✓	

Discipline Specific Elective courses (200-299)



University of Kerala

Discipline	BIOTECHNOLOGY				
Course Code	UK3DSEBIT200				
Course Title	BIOPHYSICS AND INSTRUMENTATION				
Type of Course	DSE				
Semester	III				
Academic Level	200 - 299.				
Course Details	Credit	Lecture per week	Tutorial per week	Practical per week	Total Hours/Week
	4	3 hours	0	2 hours	5
Pre-requisites	Basic Physiology, Basic Biochemistry, Fundamentals of Chemistry				
Course Summary	Biophysics and Instrumentation is an advanced graduate-level course that explores the interdisciplinary field where physics principles are applied to biological systems. The course integrates concepts from physics, biology, chemistry, and engineering to understand the physical properties of biological molecules and systems. Emphasis is placed on the development and application of various instrumentation techniques to study biological processes at molecular, cellular, and organismal levels.				

Detailed Syllabus:

Module	Unit	Content	Hrs
I	Principles of thermodynamics		6
	1	Overview of thermodynamics of biological processes –Law of thermodynamics, Gibbs free energy, energy changes in the biochemical reactions	
	2.	Electrical properties of biological compartments:-membrane potential, electrochemical gradients, ATP synthesis. (Case Study Report: 1.rotenone- proton gradient disruptor from plants 2. non-shivering thermogenesis -Thermogenin (UCP1))	
II	Biophysics of physiological events		8
	3.	Biophysics of Photosynthesis - Light reception in plants, microbes and animals. (case study:Photoreception in Bacteria: The Role of	

		Bacteriorhodopsin in Halobacterium salinarum) Resonance Energy Transfer in photosynthesis Fluorescence and phosphorescence (case study: From Glowing Jellyfish to a Revolution in Biotechnology: The Story of GFP)	
	4.	Biophysics of Vision, Muscle movements and Hearing, (case study: Ototoxicity-Induced Hearing Loss from Aminoglycoside Antibiotics), correction of vision faults, hearing aids.	
	5.	Intra and inter molecular interactions in the biological system	
III	Basic Instrumentation in Biology		19
	6.	Electrophoresis: Principle of electrophoresis, types of electrophoresis, 2-D gel electrophoresis	
	7.	Microscopy: Principle of Microscopy, various types of Microscopy - Simple, Phase contrast, Fluorescence and electron microscopy (TEM and SEM). Overview of -cryo-electron microscopy, Scanning probe microscopy and Confocal microscopy Overview of single-molecule techniques - Atomic force microscopy (AFM) (case study: Studying RNA Polymerase Transcription Using Optical Tweezers)	
	8.	Basic principles and working of: pHmeter, centrifuge, chromatography	
	9	Spectrophotometer(UV and Visible) and colorimeter - Beer-Lambert law, atomic absorption spectroscopy, IR, NMR and X-ray Crystallography and Mass Spectrometry.	
IV	Isotopes and radioisotopes		3
	10.	Application of isotopes and radioisotopes in biological research	
V	Applications of Biophysics and Instrumentation		9
	11	Drug discovery and development. Biomedical imaging and diagnostics. Biophysical approaches to understanding disease mechanisms. Emerging trends and future directions in biophysics research. Case study reports for radiotracer techniques	
	12.	Overview: computational modelling and simulation methods in biophysics. Molecular dynamics simulations for studying biomolecular structure and dynamics	

Practicals. - 30 Hrs -Essential Experiments (15 hrs), Group work (15 hrs)

Essential Experiments

1. pH Meter–Use of pH Meter, Familiarization of the instrument and Preparation Phosphate buffers and determination of pH.
2. Spectrophotometer–Familiarization of the working of the instrument, Quantitative estimation of Sugars by Dinitrosalicylic acid and Proteins by Lowry's Method
3. Development of absorption spectra of chlorophyll or any other biological sample

- Electrophoresis–demonstration of PAGE and Agarose Gel Electrophoresis

Suggested Readings

- Nelson, D. L., & Cox, M. M. (Year). *Lehninger's Biochemistry*. New York, NY: Worth Publishers.
- Voet, D., & Voet, J. G. (Year). *Biochemistry*. Boston, MA: Jones & Bartlett
- Roy, R. N. (Year). *A Textbook of Biophysics*. Calcutta, India: New Central Book Agency Pvt. Ltd.
- Nair, A. J. (Year). *Introduction to Genetic Engineering & Biotechnology*. Boston, MA: Jones & Bartlett Publishers.
- Volkenstein, M. V. (Year). *Biophysics*.
- Cantor, C. R., & Schimmel, P. R. (Year). *Biophysical Chemistry*.
- Phillips, R., Kondev, J., Theriot, J., & Garcia, H. (Year). *Physical Biology of the Cell*.
- Lakowicz, J. R. (Year). *Principles of Fluorescence Spectroscopy*.

Course Outcomes

No.	Upon completion of the course the graduate will be able to	Cognitive Level	PSO addressed
CO-1	Analyse how energy changes and conservation in the biochemical reactions is maintained	An	PSO 1, 2
CO-2	Understand the functioning of physiological events like vision, muscle movement and hearing and various types of biological interactions.	E	PSO 1
CO3	Understand basic instrumentation to analyse, elucidate and interpret a biomolecule	C	PSO 3, 4
CO4	Analyse the use of labelling of molecules in understanding the biochemical and physiological events in biology.	C	PSO 3

R-Remember, U-Understand, Ap-Apply, An-Analyse, E-Evaluate, C-Create

Note: 1 or 2 COs/module

**Name of the Course: Biophysics and instrumentation Credits: 3:0:2
(Lecture:Tutorial:Practical)**

CO No.	CO	PO/PSO	Cognitive Level	Knowledge Category	Lecture (L)/Tutorial (T)	Practical (P)
			48			

CO-1	Understand the concepts of energy conservation and changes in biological system	PSO-1,2	An	F, C	L	
CO-2	Understand the biophysics of basic physiological system	PSO1	E	P	L	
CO3	Understand Basic instrumentation to elucidate the structure of molecules	PSO3,PSO4	C	F, P	L	P
CO4	Analyse the use of labelling of molecules in understanding the biochemical and physiological events in biology.	PSO3	C	P	L	-

F-Factual, C- Conceptual, P-Procedural, M-Metacognitive

Mapping of COs with PSOs and POs :

	PSO1	PSO2	PSO3	PSO4	PSO5	PSO6	PO1	PO2	PO3	PO4	PO5	PO6
CO 1	2	3	-	-	-	-						
CO 2	3	-	-	-	-	-						
CO 3	-	-	3	4	-	-						
CO 4	-	-	2	-	-	49 -						

Correlation Levels:

Level	Correlation
-	Nil
1	Slightly / Low
2	Moderate / Medium
3	Substantial / High

Assessment Rubrics: Continuous Comprehensive Assessment:

Formative : Practical /problem/Survey/Design/Field trip based Assignment in the form of Individual or team work regarding the theory part/Follow any of the tool regarding the above practical assignment-1.Observation of practical skill/Viva/.Quiz/ In class discussions/Good Lab practices/Lab work book

Summative:Follow any of the tool - Written test (theory based)/.Open Book Test(theory based)/ Individual /team work report practical assignment/Case study report /Literature survey/Individual / Team project report oral presentation/ Laboratory report

Mapping of COs to Assessment Rubrics :

	Internal Exam	Assignment	Project Evaluation	End Semester Examinations
CO 1	✓			✓
CO 2	✓			✓
CO 3	✓			✓
CO 4		✓		✓



University of Kerala

Discipline	BIOTECHNOLOGY				
Course Code	UK3DSEBIT201				
Course Title	ENZYME ENGINEERING				
Type of Course	DSE				
Semester	III				
Academic Level	200-299.				
Course Details	Credit	Lecture per week	Tutorial per week	Practical per week	Total Hours/Week
	4	3 hours	0	2 hours	5
Pre-requisites	Biochemistry and metabolism				
Course Summary	Enzyme engineering is a multidisciplinary field that integrates principles from biology, chemistry, biochemistry, and engineering to modify enzymes for diverse applications. This course provides students with a comprehensive understanding of the basics of enzyme structure, function, and engineering methodologies. Students will explore the fundamental principles behind enzyme catalysis, techniques for enzyme characterization, and approaches for optimizing and modifying enzymes. Additionally, the course will showcase how engineered enzymes are applied in various industries including pharmaceuticals, food production, biofuels, and bioremediation.				

Detailed Syllabus:

Module	Unit	Content	Hrs
I	Introduction to enzymes		6
	1.	Enzymes, Extraction, purification, and characterization of enzymes from natural sources.	
	2.	Comparison of chemical and enzyme catalysts	
	3.	Mechanisms of enzyme action– concept of active site and energetics of enzyme-substrate complex formation, specificity of enzyme action	
II	Preparation and properties of enzymes		10
	4.	Media for enzyme production, Preparation of enzymes.	

	5.	Screening of for novel enzymes – Colorimetric, Fluorometric and biochemical assays Enzyme stabilization - stability of enzyme in various conditions, Strategies for stabilization. Enzyme specificity – level of specificity and factors affecting specificity	
	6.	Enzyme immobilization- Techniques, Benefits and applications	
III	Enzyme engineering- objectives and Methods		10
	7.	History and background of enzyme engineering, Fundamentals of protein chemistry, Objectives and Method of enzyme engineering.	
	8.	Methods- Rational design (overlap extension and whole plasmid single round PCR) and directed evolution, other methods include De novo enzyme engineering, site directed mutagenesis, random mutagenesis, DNA shuffling, phage display and mRNA display and computational methods.	
IV	Enzyme engineering applications		10
	9.	Industrial applications, such as enzyme catalysis in biocatalysis, food processing, and detergent manufacturing	
	10.	Application of enzymes in analysis -Biomedical applications, including enzyme-based therapeutics and diagnostics.	
	11.	Environmental applications, such as bioremediation and biofuel production.	
V		Computational Approaches in Enzyme Engineering Lecture Topics:	
	12.	Molecular dynamics simulations for enzyme dynamics and flexibility. Enzyme-substrate docking and binding affinity prediction. Machine learning and AI in enzyme engineering. Case studies: Computational enzyme design projects (e.g., Rosetta, FoldX).	

Practicals. - 30 Hrs -Essential Experiments (15 hrs), Group work (15 hrs)

Essential Experiments

1. Immobilize a model enzyme using different techniques.
2. Test enzyme activity before and after immobilization.
3. Evaluate factors affecting immobilization efficiency.
4. Design mutations using computational tools to improve enzyme properties (e.g., activity, specificity, stability)

- Analyze industrial enzyme formulations and their applications.

Suggested readings

- Palmer, T. (2008). *Enzymes: Biochemistry, Biotechnology, Clinical Chemistry* (2nd ed.). Affiliated East-West Press Pvt. Ltd.
- Shukla, A. (2009). *Elements of Enzymology*. Discovery Publishing House Pvt. Ltd.
- Bailey, J. E., & Ollis, D. F. (1988). *Biochemical Engineering Fundamentals* (2nd ed.). McGraw-Hill.
- Shuler, M. L., & Kargi, F. *Bioprocess Engineering Basic Concepts* (3rd ed.).

Course Outcomes

No.	Upon completion of the course the graduate will be able to	Cognitive Level	PSO addressed
CO-1	Demonstrate the comprehensive understanding of enzyme characterisation and purification including mechanism of enzyme action.	U	PSO-1,PSO3
CO-2	Analyse the preparation and screening of enzymes Evaluate enzyme immobilization	An, E	PSO3
CO3	Demonstrate historical background and objectives of enzyme engineering Investigate the methods through rational design and denovo enzyme engineering	R, An	PSO3,4
CO4	Explore the knowledge of enzyme engineering in Industry, biomedical and environmental fields	Ap	PSO4

R-Remember, U-Understand, Ap-Apply, An-Analyse, E-Evaluate, C-Create

Note: 1 or 2 COs/module

Name of the Course: Enzyme engineering Credits: 3:0:2(Lecture:Tutorial:Practical)

CO No.	CO	PO/PSO	Cognitive Level	Knowledge Category	Lecture (L)/Tutorial (T)	Practical (P)

CO-1	Evaluate the application of enzymes in various fields	PSO-1, PSO3	U	F, C	L	
CO-2	Understand the methods to improve the enzymes functions	PSO3	An, E	P	L	
CO3	Understand to preserve the structure and functions of the enzymes for long term	PSO3,4	R, An	P	L	P
CO4	Understand the application of engineered enzymes in various field	PSO4	Ap	F	L	

F-Factual, C- Conceptual, P-Procedural, M-Metacognitive

Mapping of COs with PSOs and POs :

	PSO1	PSO2	PSO3	PSO4	PSO5	PSO6	PO1	PO2	PO3	PO4	PO5	PO6
CO 1	1	-	2	-	-	-						
CO 2	-	-	3	-	-	-						
CO 3	-	-	2	2	-	-						
CO 4	-	-	-	3	-	-						

Correlation Levels:

Level	Correlation
-	Nil
1	Slightly / Low
2	Moderate / Medium
3	Substantial / High

Assessment Rubrics:

- **Continuous Comprehensive Assessment:**
- **Formative :** Practical /problem/Survey/Design/Field trip based Assignment in the form of Individual or team work regarding the theory part/Follow any of the tool regarding the above practical assignment-1.Observation of practical skill/Viva/.Quiz/ In class discussions/Good Lab practices/Lab work book
- **Summative:**Follow any of the tool - Written test (theory based)/.Open Book Test(theory based)/ Individual /team work report practical assignmen/Case study report /Literature survey/Individual / Team project report oral presentation/ Laboratory report
 - Final Exam

Mapping of COs to Assessment Rubrics :

	Internal Exam	Assignment	Project Evaluation	End Semester Examinations
CO 1	✓			✓
CO 2	✓			✓
CO 3	✓			✓
CO 4		✓		✓



University of Kerala

Discipline	BIOTECHNOLOGY				
Course Code	UK3DSEBIT202				
Course Title	INTRODUCTION TO MARINE BIOTECHNOLOGY				
Type of Course	DSE				
Semester	III				
Academic Level	200 - 299.				
Course Details	Credit	Lecture per week	Tutorial per week	Practical per week	Total Hours/Week
	4	3 hours	0	2 hours	5
Pre-requisites	Basic Biotechnology, Microbiology				
Course Summary	Marine Biotechnology is an interdisciplinary field at the forefront of scientific research, harnessing the vast potential of marine organisms and ecosystems for various applications in biomedicine, aquaculture, environmental management, and beyond. This graduate-level course delves into the principles, methodologies, and applications of marine biotechnology, equipping students with the knowledge and skills necessary to navigate the complexities of this dynamic field.				

Detailed Syllabus:

Module	Unit	Content	Hrs
I	Marine Microbial environment		6
	1	Classification of the marine environment.	
	2	Marine microbial habitats , Diversity of Marine microorganism.	
	3	Characteristics of marine microorganisms. Specialized microorganisms: Extremophiles: barophiles, thermophiles, psychrophiles, , halophiles, actinomycetes, polyextremophiles, anaerobes	
II	Techniques in Marine microbiology:		10
	4	Techniques in Marine microbiology: Sampling: Water, Sediments.	
	5	Culture based methods for isolation and, identification of microbes. define, selective and differential culture media.	
III	Bioactive molecules from marine sources		10
	6	Microbial nutrition: i) autotrophic & heterotrophic modes	
	7	Antibacterial and anti biofilm molecules produced by marine bacteria, Chitin from poriferan	

	8	Chemotaxis, Phototaxis, Bioluminescence and indicator species and Biological Rhythms	
IV	Marine bio resources		10
	9	Marine bio resources. Brief introduction - Marine microbes (viruses, bacteria, archaea, protists, fungi) Marine algae and plants (seaweeds, sea grasses, mangrove, plants) Invertebrates: sponges, cnidarians, polychaetes, crustaceans, marine worms, molluscs, echinoderms, arthropods, Non-craniate (non-vertebrate) chordates,	
	10	Molecular Adaptations in Marine Organisms: Mechanisms of adaptation to extreme environments	
V	Ecosystem functioning in marine environment		9
	11	Food web dynamics and ecosystem functioning, Microbial loop - Role of microbes in marine food web dynamics, - Biogeochemical processes: Nutrient cycling, carbon cycle, Nitrogen cycle, Sulphur cycle, Iron cycling, Phosphorus cycling and other cycles	

Practicals. - 30 Hrs -Essential Experiments (15 hrs), Group work (15 hrs)

Essential Experiments

1. Demonstrate Marine Sampling Technique- sampling marine environments (e.g., plankton nets, sediment cores, water column sampling).
2. Collection of water samples from different marine habitats (coastal, pelagic, deep-sea).
3. Isolation and characterization of marine microorganisms using culture-dependent and culture-independent techniques (e.g., metagenomics).
4. Field trip to marine ecosystems such as coral reefs, mangroves, or hydrothermal vents.
5. Identification and collection of marine organisms with potential biotechnological application

Suggested Reading

1. Blue biotechnology: production and Use of Marine Biomolecules. Stephane La Barre, Stephen S Bates. 2018. Wiley
2. Munn, C.B. , (2004) Marine Microbiology: Ecology and Applications, BIOS Scientific Publisher.
3. Krichman, D.L.,(2000), Microbial Ecology of the Oceans. Wiley-Liss, New York.
4. Paul, J.,(2001) Methods in Microbiology : marine Microbiology, Academic Press.
5. Gram, L., (2009) Microbial Spoilage of Fish and Seafood, Springer
6. Pelczar M.J. Jr., Chan E.C.S. and Kreig N.R. (2001) Microbiology, (5th Edition) CBS Publishers.

7. Josep M Gasol and David L Kirchman (2018) Marine ecology of the oceans, (3rd edition), John Wiley and Sons. Inc
8. Surajit Das Hirak Dash (2018) Microbial Diversity in the Genomic Era, Elsevier
9. Horikoshi K, Antranikian G, Bull A T, Robb F T and Stetter, K O (2011) Extremophiles Handbook, Springer
10. Madigan, Martinko, Bender, Buckley & Stahl and Thomas Brock (2017) Brock Biology of Microorganisms, Pearson

Course Outcomes

No.	Upon completion of the course the graduate will be able to	Cognitive Level	PSO addressed
CO-1	Discuss about Marine Microbial environment and its economic impact by biomaterial synthesis	U	PSO-1
CO-2	Identify the methods for biological and pharmaceutical investigation of crude extract, isolation , identification of active substances and synthesis of biomaterials	R, U,Ap	PSO3
CO3	Awareness of different bio-resources in marine environments and overview of different bioactive compounds	U,E	PSO1
CO4	Analyse different marine environments that affect overall productivity	R,U	PSO3,4

R-Remember, U-Understand, Ap-Apply, An-Analyse, E-Evaluate, C-Create
Note: 1 or 2 COs/module
Name of the Course: Introduction to marine biotechnology Credits: 3:0:2
ecture:Tutorial:Practical)

CO No.	CO	PO/PSO	Cognitive Level	Knowledge Category	Lecture (L)/Tutorial (T)	Practical (P)
CO-1	Discuss about Marine Microbial environment	PSO-1	U	F, C	L	
CO-2	Identify the methods investigation of active substances	PSO3	R, U, Ap	P	L	P

CO3	Awareness of different bio - resources in marine environment	PSO1	U, E	U	L	
CO4	Analyse different marine environment that affect overall productivity	PSO3,4	R, U	F	L	

F-Factual, C- Conceptual, P-Procedural, M-Metacognitive Mapping of COs with PSOs and POs :

	PSO1	PSO 2	PSO 3	PSO 4	PSO 5	PSO 6	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6
CO 1	2	-	-	-	-	-						
CO 2	-	-	3	-	-	-						
CO 3	-	-	1	-	-	-						
CO 4	-	-	2	3	-	-						
Correlation Levels:Level	Correlation											
-	Nil											
1	Slightly / Low											
2	Moderate / Medium											
3	Substantial / High											

Assessment Rubrics: **Continuous Comprehensive Assessment:**

Formative : Practical /problem/Survey/Design/Field trip based Assignment in the form of Individual or team work regarding the theory part/Follow any of the tool regarding the above practical assignment-1.Observation of practical skill/Viva/.Quiz/ In class discussions/Good Lab practices/Lab work book

Summative: Follow any of the tool - Written test (theory based)/.Open Book Test(theory based)/ Individual /team work report practical assignmen/Case study report /Literature survey/Individual / Team project report oral presentation/ Laboratory report

Mapping of COs to Assessment Rubrics :

	Internal Exam	Assignment	Project Evaluation	End Semester Examinations
CO 1	✓			✓
CO 2	✓			✓
CO 3	✓			✓
CO 4		✓		✓



University of Kerala

Discipline	BIOTECHNOLOGY				
Course Code	UK3DSEBIT203				
Course Title	BIOMOLECULAR INTERACTIONS & CELL SIGNALLING				
Type of Course	DSC				
Semester	III				
Academic Level	200 - 299.				
Course Details	Credit	Lecture per week	Tutorial per week	Practical per week	Total Hours/Week
	4	3 hours	0	2 hours	5
Pre-requisites	Cell biology & molecular biology				
Course Summary	This course covers a wide range of topics related to host-parasite interaction, cellular communication, and signalling regulation, providing students with a comprehensive understanding of these complex biological processes				

Detailed Syllabus:

Module	Unit	Content	Hrs
I	Introduction to Biomolecular Interactions		6
	1	Overview of biomolecules: proteins, nucleic acids, carbohydrates, and lipids Forces governing biomolecular interactions: electrostatic, hydrophobic, hydrogen bonding Techniques for studying biomolecular interactions: spectroscopy, chromatography, mass spectrometry	
	2	Cellular interactions: Extracellular matrix molecular interaction, matrix proteins, integrins, focal adhesions and hemidesmosomes; Cell-cell interactions and cellular junctions	
II	General principles of cell communication		10
	3	Basics of cell signaling: autocrine, paracrine, endocrine signaling Signal transduction pathways: receptor activation, intracellular signaling cascades- Hormones and their receptors, cell surface receptor, Ser/Thr protein kinases and phosphatases, Tyr phosphorylation signaling receptor and non-receptor TKs, Protein Kinase (PKC) Signaling, cytokine receptors and the JAK-STAT Pathway signaling through Gprotein coupled receptors, Toll like Receptors, Inflammasomes , Role of NO as an Intercellular Messenger , Regulation of cell signaling: feedback mechanisms, cross-talk between	

		pathways	
	4	general principles of cell communication - cell adhesion and roles of different adhesion molecules, gap junctions, extracellular matrix, integrins, neurotransmission and its regulation	
	5	Host parasite interaction: Recognition and entry processes of different pathogens like bacteria, viruses into host cells, alteration of host cell behaviour by pathogens, bacterial signalling systems, bacterial chemotaxis and quorum sensing.	
III	Protein-Protein Interactions		10
	6	Structural basis of protein-protein interactions	
	7	Methods for studying protein-protein interactions: yeast two-hybrid, co-immunoprecipitation, surface plasmon resonance	
	8	Applications in drug discovery and protein engineering, Post-translational modifications and their impact on protein function Membrane protein interactions and their significance in cellular processes	
IV	Protein-DNA Interactions		10
	9	Mechanisms of protein-DNA recognition and binding	
	10	DNA-binding domains and transcription factors	
	11	Genome-wide approaches for studying protein-DNA interactions: ChIP-seq, DNase-seq	
V	Cell Signaling in Disease		9
	12	Aberrant signaling in cancer, neurodegenerative diseases, and metabolic disorders Targeting signaling pathways for therapeutic interventions Case studies illustrating the role of biomolecular interactions in disease progression and treatment	

Practicals. - 30 Hrs -Essential Experiments (15 hrs), Group work (15 hrs)

Essential Experiments

1. Stimulate cells with a signaling molecule (e.g., growth factor or hormone).
2. Monitor the activation of downstream signaling components using techniques like immunoblotting, immunofluorescence, or ELISA.
3. Examine the effect of signaling pathways on gene expression.
4. Treat cells with pathway activators or inhibitors.
5. Measure changes in mRNA expression using qRT-PCR, RNA-seq, or microarray analysis.

Suggested Reading

1. G. Karp, Cell and Molecular Biology, 5th Edn., Wiley, 2007
2. D. L. Wheeler, Y. Yarden, Receptor Tyrosine Kinases: Structure, Functions and Role in Human Disease, Springer, 2014

3. Q. A. Acton, Receptor Protein-Tyrosine Kinases: Advances in Research and Application, ScholarlyEditions, 2012
4. B. Alberts, A. Johnson, J. Lewis, and M. Raff, Molecular Biology of the Cell, 5th Edn., Garland Science, 2008.
5. H. Lodish, A. Berk, C.A. Kaiser, and M. Krieger, Molecular Cell Biology, 6th Edn., W. H. Freeman, 2007.

Course Outcomes

No.	Upon completion of the course the graduate will be able to	Cognitive Level	PSO addressed
CO-1	Explore the role of biomolecular interactions in cellular processes and disease mechanisms.	U	PSO-1,2
CO-2	Explain the fundamental principles of cell communication, including signaling pathways, signal transduction, and cellular responses to extracellular stimuli.	R, U	PSO1
CO-3	Analyze various cell signaling pathways, including those mediated by hormones, growth factors, and cytokines, and their corresponding receptors.	U,AN	PSO1,PSO3,4
CO-4	Explore the role of biomolecular interactions in cellular processes and disease mechanisms..	U,AN	PSO4

R-Remember, U-Understand, Ap-Apply, An-Analyse, E-Evaluate, C-Create *Note: 1 or 2 COs/module*

Name of the Course: Biomolecular interactions & cell signalling Credits: 3:0:2
cture:Tutorial:Practical)

CO No.	CO	PO/PSO	Cognitive Level	Knowledge Category	Lecture (L)/Tutorial (T)	Practical (P)
CO-1	Understand basics of host –pathogen interaction	PSO-1,2	U	F, C	L	-
CO-2	Explain fundamentals of cell signalling	PSO1	R, U	P	L	-
CO-3	Discuss various receptors and signalling molecules	PSO1,PSO3,4	U,AN	F	L	-

CO-4	Analyse the cell signalling regulation	PSO4	U,AN	C	L	-
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F-Factual, C- Conceptual, P-Procedural, M-Metacognitive

Mapping of COs with PSOs and POs :

	PSO1	PSO2	PSO3	PSO4	PSO5	PSO6	PO1	PO2	PO3	PO4	PO5	PO6
CO 1	2	2	-	-	-	-						
CO 2	2	-	-	-	-	-						
CO 3	2	-	2	3	-	-						
CO 4	-	-	-	3	-	-						

Correlation Levels:

Level	Correlation
-	Nil
1	Slightly / Low
2	Moderate / Medium
3	Substantial / High

Assessment Rubrics: **Continuous Comprehensive Assessment:**

Formative : Practical /problem/Survey/Design/Field trip based Assignment in the form of Individual or team work regarding the theory part/Follow any of the tool regarding the above practical assignment-1.Observation of practical skill/Viva/.Quiz/ In class discussions/Good Lab practices/Lab work book

Summative:Follow any of the tool - Written test (theory based)/.Open Book Test(theory based)/ Individual /team work report practical assignmen/Case study report /Literature survey/Individual / Team project report oral presentation/ Laboratory report

Mapping of COs to Assessment Rubrics :

	Internal Exam	Assignment	Project Evaluation	End Semester Examinations
CO 1	✓		64	✓

CO 2	✓			✓
CO 3	✓			✓
CO 4		✓		✓

Value Added Courses (200-299)



Course Title	IPR,BIOETHICS AND BIOSAFETY				
Type of Course	VAC				
Semester	III				
Academic Level	200 - 299 .				
Course Details	Credit	Lecture per week	Tutorial per week	Practical per week	Total Hours/Week
	3	2 hours	1		3
Pre-requisites	Essentials of Biotechnology				
Course Summary	This course provides an in-depth exploration of the intersection of intellectual property rights, bioethics, and biosafety within the context of modern scientific research and biotechnology. It aims to equip students with a comprehensive understanding of the legal, ethical, and safety considerations inherent in the development, dissemination, and regulation of biotechnological innovations.				

Detailed Syllabus:

Module	Unit	Content	Hrs
I	Basics of Intellectual Property Rights		9
	1	Types of IPR- Industrial – Patent (Patentable and Non-patentable, Types of Patent, Patenting life), Design, Trademark, Trade secret, Geographical Indication; Non industrial – Copyright (Types, exclusions), Rights of traditional knowledge holders, Peoples biodiversity register	
	2	Legal protection of Biotechnological Inventions, Pros & Cons of IPR; World Intellectual Property Right Organization (WIPO), TRIPS agreement, UPOV Convention; Patent Infringement (Case Study), Plagiarism, Plagiarism Detection Softwares and ways to avoid plagiarism	
II	Biosafety		9
	3	Biosafety Issues in Biotechnology, Biosafety-Different levels of Biosafety, Biosafety levels of specific Microorganisms (e.g. Non-pathogenic E. coli, HIV, Staphylococcus aureus, West Nile virus, Tuberculosis, Ebola); Biological Safety Cabinets; Biocontainment – Definition and Types (Biological and Physical)	

	4	Principles and practices of biosafety in laboratory and industrial settings; Risk assessment methodologies and strategies for mitigating biological hazards (A brief account of the steps involved)	
III	Guidelines of Biosafety		9
	5	Biosafety guidelines and regulations (National and International) – Brief account on the guidelines of WHO, DBT GoI, Cartagena Protocol; GMOs and LMOs – Definition	
	6	Institutional Biosafety Committee (IBSC), Review Committee on Genetic Manipulation (RCGM), Genetic Engineering Appraisal Committee (GEAC), for GMO applications	
	7	Assessment and management of risks associated with GMO (Overview)	
IV	Bioethics		9
	8	Bioethics-Introduction, Key Concepts and Principles of Bioethics - Autonomy, Beneficence, Non-maleficence, Justice. Concepts of Animal Ethics, Animal ethical theories - Animal Rights (Deontology) and Utilitarianism	
	9	Biotechnology and Ethics; Ethical issues related to research in embryonic stem cells and cloning; Ethical, Legal and Social Implications (ELSI) of Human Genome Project; Ethical issues related to animal experiments	
V	Essentials of scientific experiments		9
	10	Values in science, Misconduct in science, Negligence and error, Conflict of interest	
	11	Analyze case studies and real-world examples – to understand the practical application of IPR in biotechnology, Protection of Plant Varieties and Farmer's Rights	
	12	Debate contemporary bioethical issues such as gene editing, and access to healthcare	

Suggested Reading

1. Intellectual Property Rights: A Practical Guide to Content, Protection, and Exploitation" by Stephen Johnson
2. "Intellectual Property: A Very Short Introduction" by Siva Vaidyanathan
3. "Intellectual Property in the New Technological Age" by Robert P. Merges and Peter S. Menell
4. Bioethics: Principles, Issues, and Cases" by Lewis Vaughn
5. "Principles of Biomedical Ethics" by Tom L. Beauchamp and James F. Childress
6. "Bioethics: An Introduction" by Marianne Talbot
7. Biosafety in Microbiological and Biomedical Laboratories" by Centers for Disease Control and Prevention (CDC) and National Institutes of Health (NIH)
8. "Biosafety in Industrial Biotechnology" by Preeti Jain and Rakesh Singh
9. "Handbook of Laboratory Health and Safety" by Robert H. Hill Jr. and David W. Smith

Course Outcomes

No.	Upon completion of the course the graduate will be able to	Cognitive Level	PSO addressed
CO1	Able to recognize the importance of Intellectual Property Rights and distinguish various forms of IPR	An	PSO-2,5
CO2	Awareness about legal protection of Intellectual Property and empower students to protect their Intellectual Property	C	PSO-2,5
CO3	Gain knowledge of Biosafety, associated risks and containment strategies	An	PSO -5
CO4	Get an insight into Biosafety guidelines and regulations with specific emphasis to Genetically Modified Organisms	E	PSO -5
CO5	Understand the basics of Bioethics and relevance of Ethical issues related to research in different areas of Biotechnology and present arguments related to bioethical issues in a logical manner	C	PSO-2

R-Remember, U-Understand, Ap-Apply, An-Analyse, E-Evaluate, C-Create

Note: 1 or 2 COs/module

Name of the Course: IPR, bioethics and biosafety Credits: 2:1:0 (Lecture:Tutorial:Practical)

CO No.	CO	PO/PSO	Cognitive Level	Knowledge Category	Lecture (L)/Tutorial (T)	Practical (P)
CO1	Able to recognize the importance of Intellectual Property Rights and distinguish various forms of IPR	PSO - 2, 5	An	F,C	L	
CO2	Awareness about legal protection of Intellectual Property and empower students to protect their Intellectual Property	PSO - 2, 5	C	P, M	L	
CO3	Gain knowledge of Biosafety, associated risks and containment strategies	PSO - 5	An 68	F	L	

CO4	Get an insight into Biosafety guidelines and regulations with specific emphasis to Genetically Modified Organisms	PSO - 5	E	C	L	
CO5	Understand the basics of Bioethics and relevance of Ethical issues related to research in different areas of Biotechnology and present arguments related to bioethical issues in a logical manner	PSO - 2	C	C, M	L	

F-Factual,C-Conceptual,P-Procedural,M-Metacognitive

Mapping of COs with PSOs and POs :

	PSO1	PSO2	PSO3	PSO4	PSO5	PSO6	PO1	PO2	PO3	PO4	PO5	PO6
CO1	-	2	-	-	3	-						
CO2	-	3	-	-	3	-						
CO3	-	-	-	-	2	-						
CO4	-	-	-	-	3	-						
CO5	-	3	-	-	-	-						

Correlation Levels:

Level	Correlation
-	Nil
1	Slightly/Low

2	Moderate/ Medium
3	Substantial/High

Assessment Rubrics: **Continuous Comprehensive Assessment:**

Formative : Practical /problem/Survey/Design/Field trip based Assignment in the form of Individual or team work regarding the theory part/Follow any of the tool regarding the above practical assignment-1.Observation of practical skill/Viva/.Quiz/ In class discussions/Good Lab practices/Lab work book

Summative:Follow any of the tool - Written test (theory based)/.Open Book Test(theory based)/ Individual /team work report practical assignmen/Case study report /Literature survey/Individual / Team project report oral presentation/ Laboratory report

FinalExam

MappingofCOstoAssessmentRubrics:

	InternalExam	Assignment	ProjectEvaluation	EndSemester Examinations
CO1	✓	✓		✓
CO2	✓			✓
CO3	✓			✓
CO4	✓	✓		✓
CO 5	✓	✓		✓

